



Cambridge International AS & A Level

CANDIDATE
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BIOLOGY

9700/31

Paper 3 Advanced Practical Skills 1

October/November 2022

2 hours

You must answer on the question paper.

You will need: The materials and apparatus listed in the confidential instructions

INSTRUCTIONS

- Answer **all** questions.
- Use a black or dark blue pen. You may use an HB pencil for any diagrams or graphs.
- Write your name, centre number and candidate number in the boxes at the top of the page.
- Write your answer to each question in the space provided.
- Do **not** use an erasable pen or correction fluid.
- Do **not** write on any bar codes.
- You may use a calculator.
- You should show all your working and use appropriate units.

INFORMATION

- The total mark for this paper is 40.
- The number of marks for each question or part question is shown in brackets [].

For Examiner's Use	
1	
2	
Total	

This document has **16** pages. Any blank pages are indicated.



- 1 Some fruit contains large quantities of ascorbic acid (vitamin C). Ascorbic acid can be absorbed through the partially permeable wall of the gut.

You will investigate the rate at which ascorbic acid from a 2 g dm^{-3} ascorbic acid solution diffuses across a partially permeable membrane. Dialysis (Visking) tubing acts as a partially permeable membrane.

You are provided with the materials shown in Table 1.1.

Table 1.1

labelled	contents	hazard	quantity
A	2 g dm^{-3} ascorbic acid solution	irritant	80 cm^3
W	distilled water	none	200 cm^3
D	dialysis tubing in distilled water	none	20 cm
I	iodine solution	irritant	25 cm^3
S	starch solution	none	20 cm^3

It is recommended that you wear suitable eye protection.

You will need to:

- allow ascorbic acid to diffuse out of the dialysis tubing into the distilled water surrounding the dialysis tubing
- estimate the concentration of ascorbic acid that has diffused out of the dialysis tubing.

Carry out step 1 to step 27.

- step 1 Tie a knot in the dialysis tubing as close as possible to one end, so that the end is sealed.
- step 2 To open the other end, wet the dialysis tubing and rub the tubing gently between your fingers and thumb.
- step 3 Put 10 cm^3 of ascorbic acid solution **A** into the open end of the dialysis tubing.
- step 4 Rinse the outside of the dialysis tubing by dipping it in the container of water labelled **D**.
- step 5 Carefully place the filled dialysis tubing bag into the large test-tube and secure in position using an elastic band, as shown in Fig. 1.1.

3

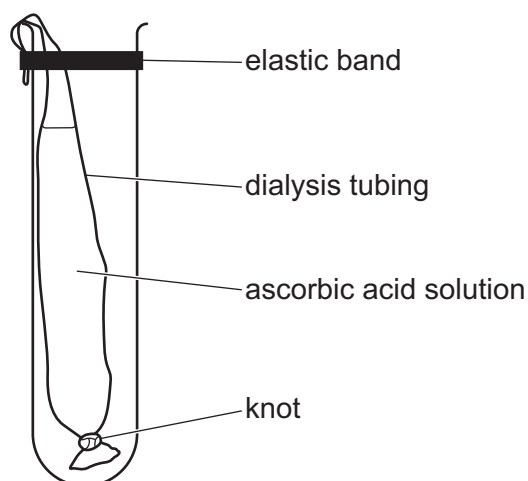


Fig. 1.1

You will need to fill the large test-tube with a known volume of distilled water so that it just covers the liquid in the dialysis tubing.

step 6 Use a syringe to add the distilled water to the large test-tube and record in **(a)(i)** the volume you added.

(a) (i) State the volume of distilled water you added.

volume of water cm³ [1]

step 7 Start timing and leave the dialysis tubing bag in the distilled water for at least 15 minutes.

While you are waiting continue with this question.

You will need to dilute the 2 g dm⁻³ ascorbic acid solution, **A**, to provide a range of known concentrations.

You will need to make up 20 cm³ of each concentration of ascorbic acid.

Table 1.2 shows how to make up two of the concentrations you should use.

(ii) Decide which other concentrations of ascorbic acid to make and complete Table 1.2.

Table 1.2

concentration of ascorbic acid /g dm ⁻³	volume of ascorbic acid solution /cm ³	volume of distilled water /cm ³
0.0	0.0	20.0
2.0	20.0	0.0

[2]

step 8 Using the beakers provided, make up the concentrations of ascorbic acid stated in Table 1.2.

step 9 Put 1 cm³ of starch solution **S** into a clean test-tube.

step 10 Put 5 cm³ of the 0 g dm⁻³ ascorbic acid solution (distilled water) into the same test-tube. Shake gently to mix.

step 11 Fill a syringe with 2 cm³ iodine solution **I**.

step 12 Add one drop of **I** to the mixture of **S** and ascorbic acid solution, as shown in Fig. 1.2. Mix gently.

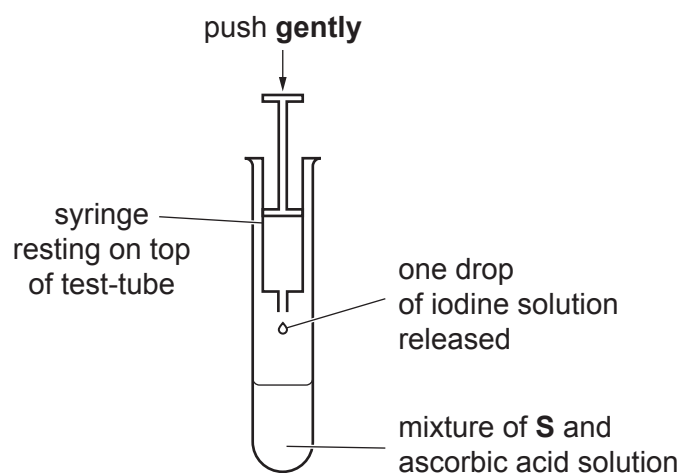


Fig. 1.2

5

- step 13 You should see a blue colour. This is the end-point you are looking for when you test the remaining ascorbic acid solutions (step 21).
- step 14 Record the volume of iodine solution you have added.
- step 15 Put 1 cm³ of starch solution **S** into a clean test-tube.
- step 16 Put 5 cm³ of the lowest concentration of the remaining ascorbic acid solutions into the same test-tube. Shake gently to mix.
- step 17 Fill the syringe containing iodine solution **I** to the 2 cm³ level again.
- step 18 Add one drop of **I** to the mixture of **S** and ascorbic acid solution as shown in Fig. 1.2. Mix well.
- step 19 Continue adding drops one at a time, mixing after each drop, until you see a blue colour.
- step 20 As soon as you see a blue colour, wait 10 seconds. If the blue colour disappears then add another drop.
- step 21 Continue adding drops until the mixture stays blue for at least 10 seconds. This is the end-point.
- step 22 Record the volume of iodine solution you have added.
- step 23 Repeat step 15 to step 22 with the other concentrations of ascorbic acid you prepared in step 8.

Record your results in **(a)(iii)**.

- (iii)** Record your results in an appropriate table.

[5]

step 24 Remove the dialysis tubing from the large test-tube and place it in the beaker labelled **For waste**. Stop timing and record the time that the dialysis tubing has been left in the distilled water (time from step 7 to step 24).

time =

step 25 Pour the solution from the large test-tube into a small beaker.

step 26 Use a syringe to remove a 5 cm^3 sample of the solution from the small beaker.

step 27 Repeat step 15 to step 22 using this sample. Record your result in **(a)(iv)**.

(iv) Record the volume of iodine added. [1]

(v) Use your results in **(a)(iii)** and **(a)(iv)** to estimate the concentration of ascorbic acid in the solution outside the dialysis tubing bag.

concentration of ascorbic acid [1]

(vi) Calculate the **rate** of diffusion of ascorbic acid out of the dialysis tubing.

Show your working.

rate of diffusion [2]

(vii) Identify **two** significant sources of error when finding the concentration of ascorbic acid in the sample from the large test-tube.

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..... [2]

(viii) Suggest how you would make **two** improvements to this investigation.

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..... [2]

- (b) Fruit can be preserved by reducing its water content. This can be done by cutting the fruit into small cubes and soaking the cubes in concentrated sucrose solution.

Cubes of melon were soaked in 45% sucrose solution for 20 hours at a temperature of 25 °C.

The water content of the melon was determined every 4 hours.

The results are shown in Table 1.3.

Table 1.3

time / hours	water content /g of water for each g of melon
0	1.44
4	0.83
8	0.42
12	0.25
16	0.20
20	0.20

(i) Plot a graph of the data in Table 1.3 on the grid in Fig. 1.3.

Use a sharp pencil.

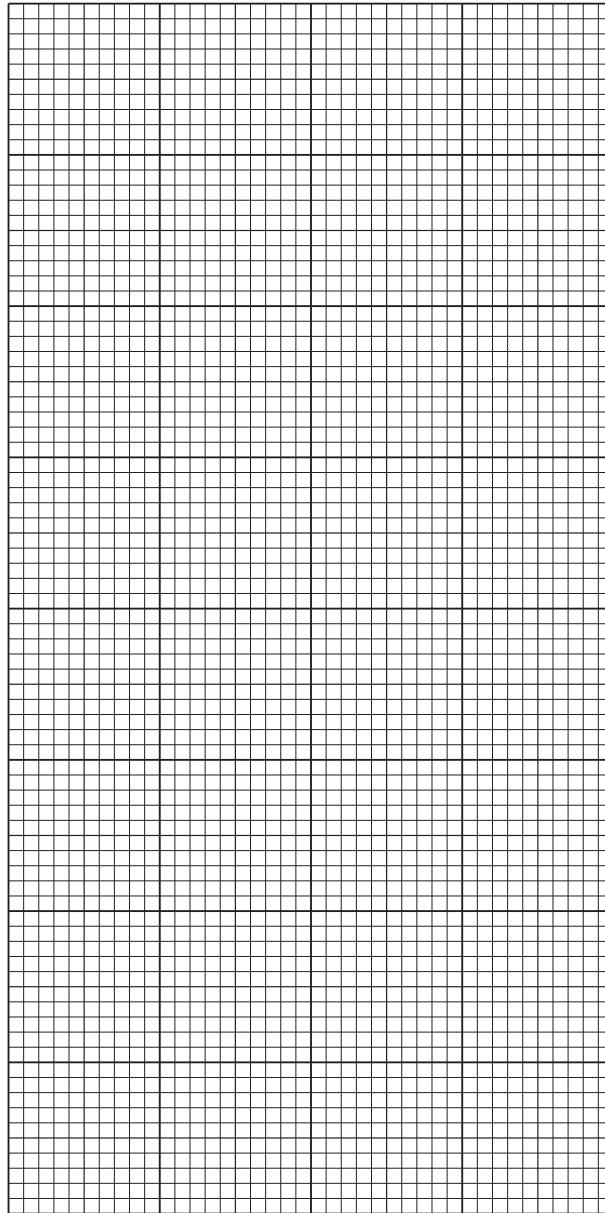


Fig. 1.3

[4]

(ii) Explain the change in water content of the melon cubes between 16 hours and 20 hours.

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..... [2]

2 **J1** is a slide of a stained transverse section through a plant stem.

- (a) (i) Draw a large plan diagram of the region of the stem on **J1** indicated by the shaded region in Fig. 2.1. Use a sharp pencil.

Use **one** ruled label line and label to identify the epidermis.

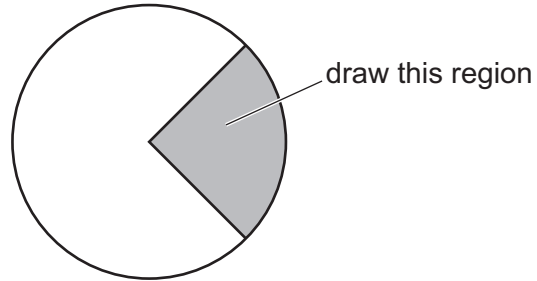


Fig. 2.1

[5]

(ii) Observe the xylem vessel elements of the section on **J1**.

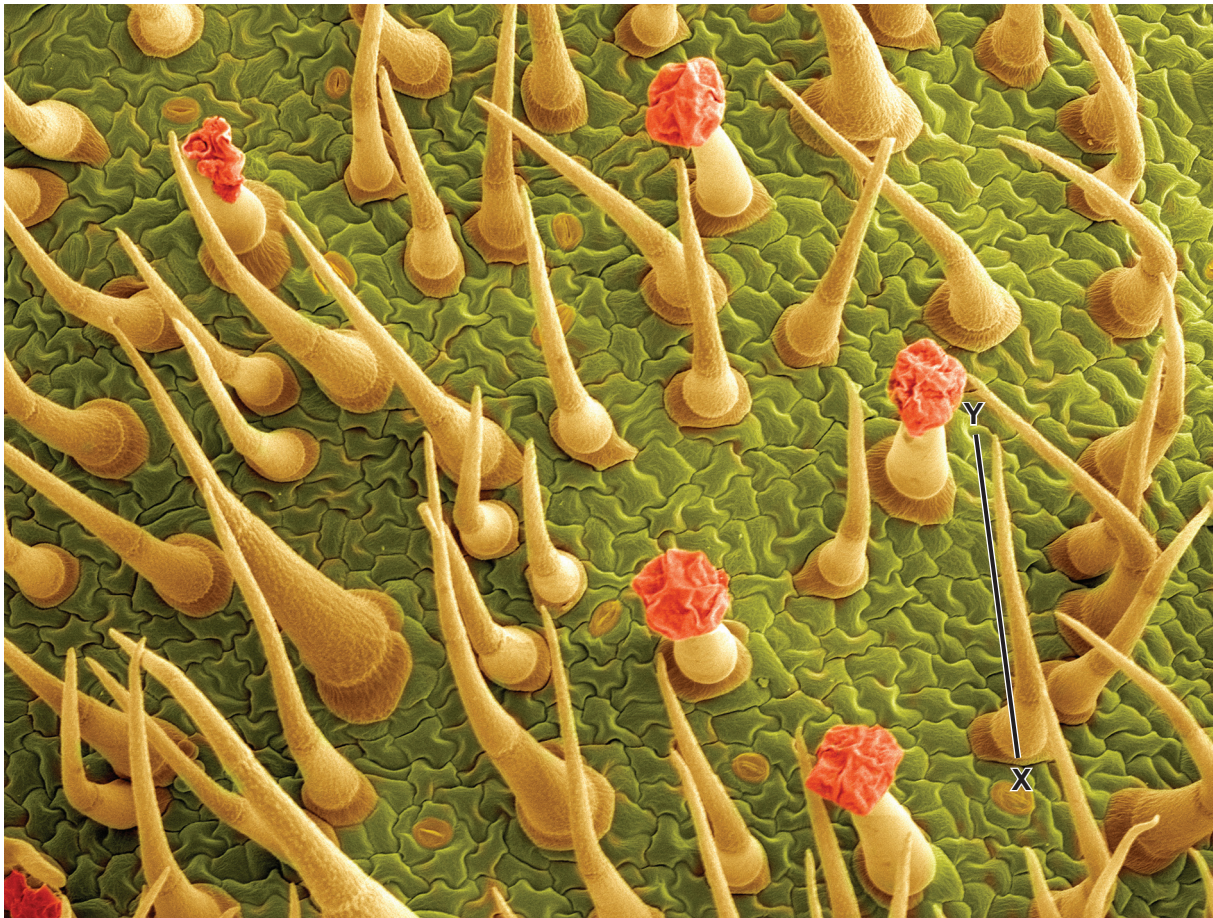
Select **four** adjacent xylem vessel elements.

Each xylem vessel element must touch at least one other xylem vessel element.

- Make a large drawing of these **four** xylem vessel elements.
- Use **one** ruled label line and label to identify the cell wall of **one** xylem vessel element.

[5]

(b) Fig. 2.2 is a scanning electron micrograph of trichomes on the surface of a leaf.




scale bar

 100 μm

Fig. 2.2

- (i) Use the scale bar and line X–Y on Fig. 2.2 to calculate the actual length of one trichome. Show your working and use appropriate units.

..... [3]

- (ii) Add a labelled line to Fig. 2.2 to identify a stoma. [1]

(iii) Fig. 2.3 is a scanning electron micrograph of trichomes on the leaf of a different plant.

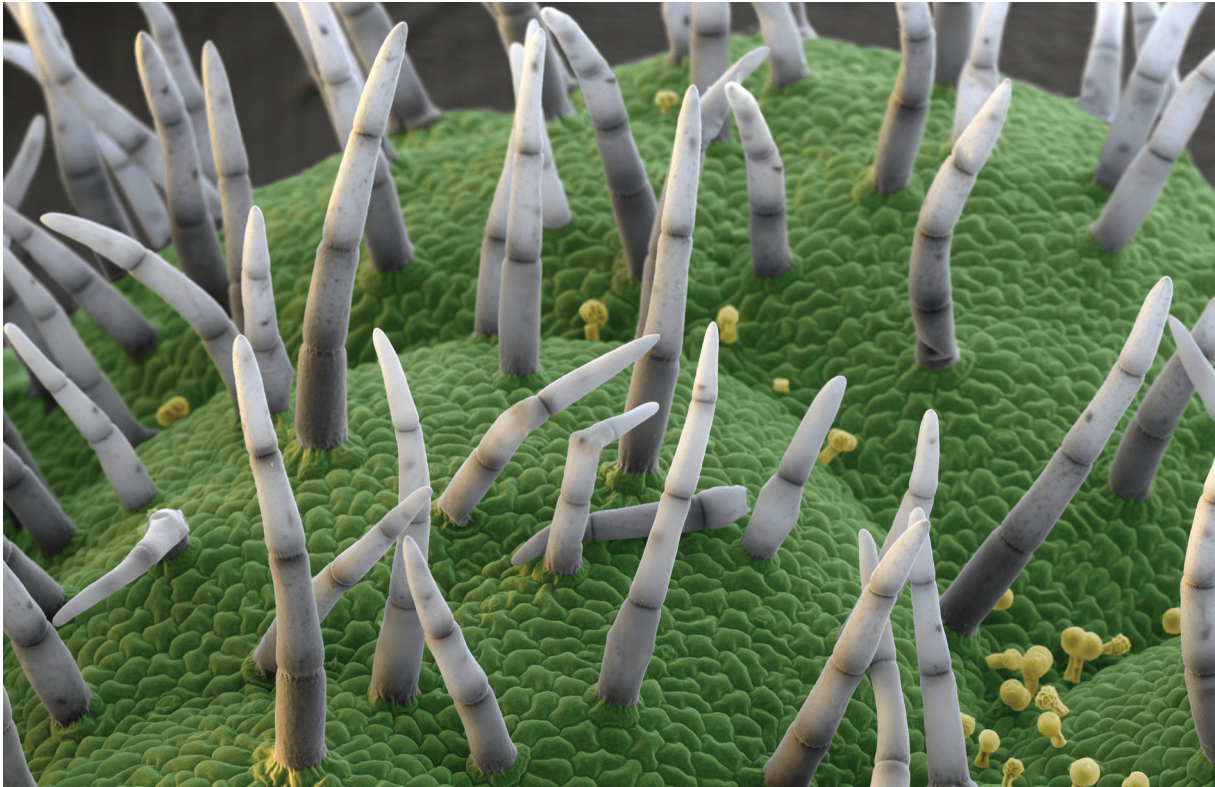


Fig. 2.3

Observe the trichomes in Fig. 2.2 **and** Fig. 2.3.

Identify **one** observable similarity and **two** observable differences, other than size and colour, between the trichomes in Fig. 2.2 and the trichomes in Fig. 2.3.

Record these observations in an appropriate table.

[4]

[Total: 18]

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